

Project: Guidance development and case study documentation of green chemistry and technologies



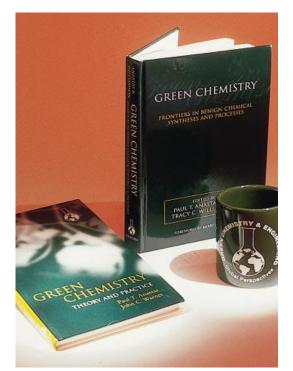
MICROORGANISMS IN GREEN CHEMISTRY

Dr. Vladimir P. Beškoski, Associated Proffessor University of Belgrade - Faculty of Chemistry

University of Belgrade – Faculty of Chemistry, 30.11.2018 https://greenchemistryserb.wixsite.com/green-serbia

WHAT IS GREEN CHEMISTRY?

"Green chemistry is the design of new products and processes that reduce or eliminate the use and generation of hazardous substances"



Paul Anastas 1997



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Presidential Green Chemistry Challenge Winners

On this page:

Award winners by year with links to technology summaries and podcasts (for some).

On other pages:

- Summaries of all winning technologies in PDF format: 1996-2016 PGCC Award Recipients Booklet
- Winning technologies indexed by technology
- · Winning technologies indexed by industry sector

Disclaimer: Mention of trade names, products, or services does not convey official EPA approval, endorsement, or recommendation.

https://www.epa.gov/greenchemistry/presidential-greenchemistry-challenge-winners

RECALL WHAT IS THE BIOTECHNOLOGY!

»Biotechnology is the integration of natural sciences and engineering sciences in order to achieve the application of organisms, cells, parts thereof and molecular analogues for products and services»

(General Assembly of the European Federation of

Biotechnology, 1989)

»Biotechnology is the application of science and technology to living organisms as well as parts, products and models thereof, to alter living or non-living materials for the production of knowledge, goods and services» (OECD, 2002)

Color Type Health, Medical, Diagnostics Red Yellow Food Biotechnology, Nutrition Science Blue Aquaculture, Coastal and Marine Biotech Agricultural, Environmental Biotechnology - Biofuels, Biofertilizers, Green Bioremediation, Geomicrobiology Arid Zone and Desert Biotechnology Brown Dark Bioterrorism, Biowarfare, Biocrimes, Anticrop warfare Patents, Publications, Inventions, IPRs Purple White Gene-based Bioindustries Gold Bioinformatics, Nanobiotechnology Classical Fermentation and Bioprocess Technology Grey

Area of Biotech Activities

http://www.ejbiotechnology.info/index.php/ejbiotechnology/article/view/1114/1496

Biotechnology meets some of the principles of green chemistry and environmental protection:

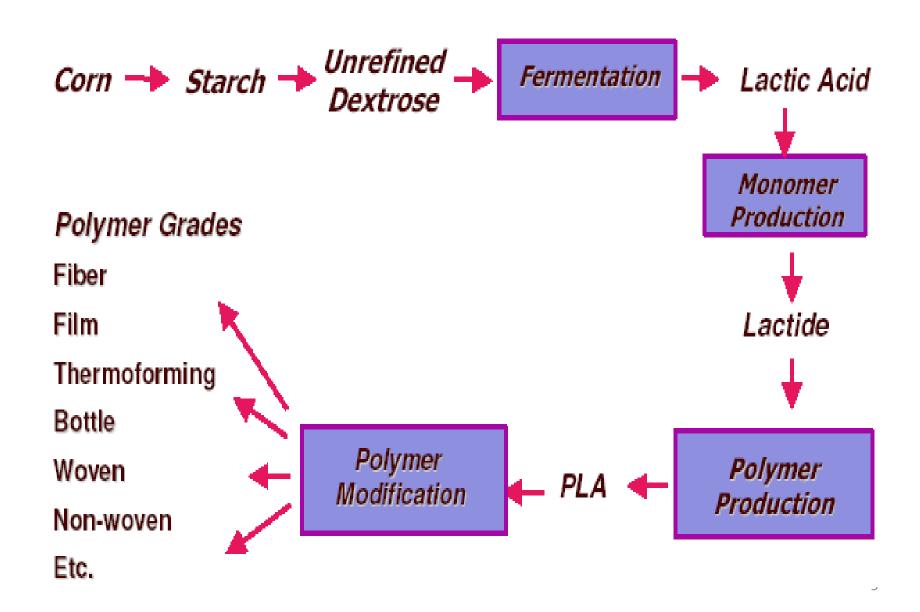
- > mild reaction conditions (physiological pH and temperature)
- use of enzymes, microorganisms and plant tissues;
- environmentally compatible catalysts and solvents (often water) in combination with high activity and selectivity in multifunctional molecules;
- sustainable approaches for the production of fine chemicals (aromas, polymers, pharmaceuticals and enzymes);
- use of renewable sources or agroindustrial residues;
- recycling of biocatalysts;
- > finally minimizing of waste.

Some of the interesting examples of biotechnology - green chemistry are fermentation processes that produce:

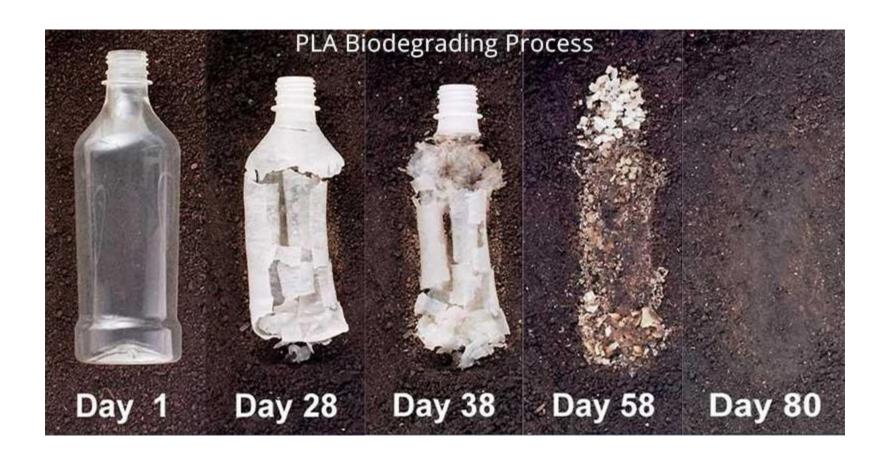
- biodegradable polyhydroxyalkanoate and polylactate "natural plastic" from renewable sources
- rhamnolipids, <u>biosurfactants</u> that can replace synthetic surfactants and apply in the petroleum industry, agriculture, personal care products ...;
- enzymes for improving recycled paper processing;
- plastics based on greenhouse gases, such as methane;
- <u>simvastatin</u>, the medicine for the treatment of high cholesterol (Aspergillus terreus);
- **>** ...

Biopolymers

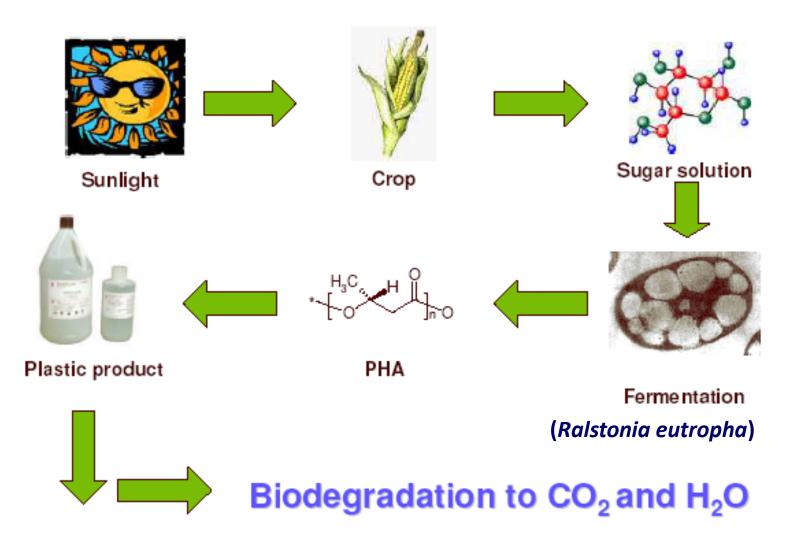
Bioplastics - Polylactic acid - PLA for production of plastics



Bioplastics - Polylactic acid - PLA for production of plastics

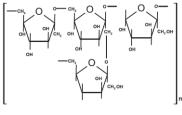


Bioplastics – Polyhydroxyalkanoates PHA's

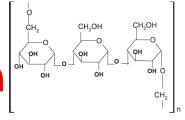


In 2016 260 million tons of plastic were produced worldwide and around 4,2 million tons of bioplastics. Estimations are that 6,1 million tons will be produced in 2021.

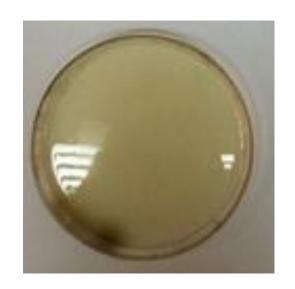
PHA 3-5 EUR/kg Propilen 1 EUR/kg



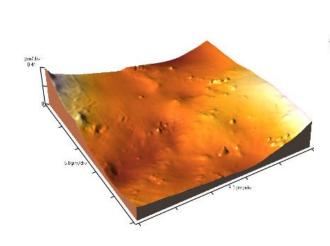
Bioplastics – levan & pullulan

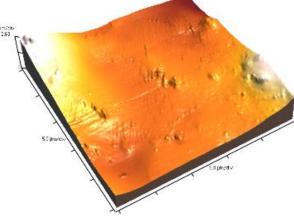


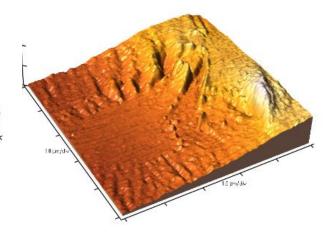
















NASLOVNA

VESTI

STARS

SPORT

HRONIKA

REGION

PLANETA

ZABAVA

EXTRA

STIL





A MASOVNA TUČA! Sevale su pesnice, prštale uvrede, a JEZIVE SCENE su obišle ceo svet (VIDEO)

DAČIĆ: Srbiji nisu potrebne verb NAJNOVIJE VESTI >





PENZIONERI, 3,000 DINARA STIŽE VAM lednokratna pomoć



ŠOK MODA U JUŽNOM DELU MITROVICE: I lutke butiku obukli u



RAMINA VELIKOALBANSKA PROVOKACIJA:



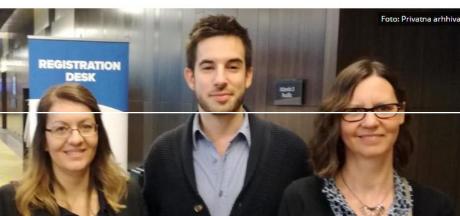
NJUJORK TAJMS NAHVALIO NIŠKI MEGAPROIEKAT: Umesto kasarne.











PRVI PUT U SRBIJI EKO-**INOVATIVNO RECIKLIRANJE:** Bakterije koje jedu plastiku da bi proizvele bioplastiku!



NAJNOVIJE

NAJČITANIJE

REAKCIJE

SVAKOG JUTRA JELA SAM OVSENU KAŠU SA TOPLOM VODOM: Kada sam videla rezultate 30 dana posle, **ŠOKIRALA SAM SE!**

GRUZIJA IZABRALA PRVU ŽENU PREDSEDNIKA: Salome Zurabišvili pobedila u drugom krugu predsedničkih izbora 6 11

29. NOVEMBAR: Dan kada je rođena Jugoslavija! Q 216 R 383

TAČI ZA GLAS AMERIKE: Puna mu usta razgraničenja i promene rezolucije 1244, a za takse pravi se lud! Pa još Beogradu postavlja i ovaj uslov?!

VAJLDER UBO FJURIJA PRSTOM U OKO, A ONDA JE NASTALA MASOVNA TUČA! Sevale su pesnice, prštale uvrede, a JEZIVE SCENE su obišle ceo svet (VIDEO)

Biosynthesis and recent advances in the production of bacterial cellulose

Cellulose is the most common natural polymer worldwide and 10¹⁴ t of cellulose pulp are produced each year. Due to the presence of components other than cellulose, different chemical treatments employing highly polluting chemical products, such as chlorine gas, caustic soda, carbon disulfide, carbon monoxide, and carbon dioxide, are often needed prior to use in order to obtain pure cellulose.

Donini et al. compared the productivity of cellulose from plants and microorganisms in order to determine the advantages of producing cellulose from microorganisms. They compared the **production of cellulose** from **1 ha of eucalyptus** with a mean annual increment MAI of 25 t/ha/year. With 7 years from planting to cultivation, yielding about 45 % cellulose contents, this process would yield **about 80 t of cellulose/ha after 7 years of cultivation**.

The same production could be achieved with bacteria to a hypothetical yield of 15 g/L in 50 h of culture (average of 0.3 g/h) in a bioreactor of 500 m³ in approximately 22 days.

Unlike plant cellulose, BNC is produced in the pure form, devoid of lignin, hemicellulose, pectin, or any other compound present in the plant pulp!!!

Bacterial nanocellulose - BNC

BNC can provide improved mechanical qualities to the biomaterial owing to its biocompatibility, biofunctionality, lack of toxicity, and ease of sterilization.

BNC is a highly crystalline **linear polymer of glucose** synthesized mainly by the bacterium *Gluconacetobacter xylinus* (formerly named *Acetobacter xylinus*). Although BNC production has been studied primarily in *G. xylinus*, other microorganisms also exhibit the ability to synthesize this biopolymer, such as other species of *Gluconacetobacter*, *Agrobacterium tumefaciens*, *Rhizobium* spp., and Gram positive *Sarcina ventriculli*.

One important aspect in BNC production is **to identify a low-cost culture medium** that can
improve the yield of BNC and be used as an
economically viable solution for application in a
range of fields.



Bacterial nanocellulose – Carbon source

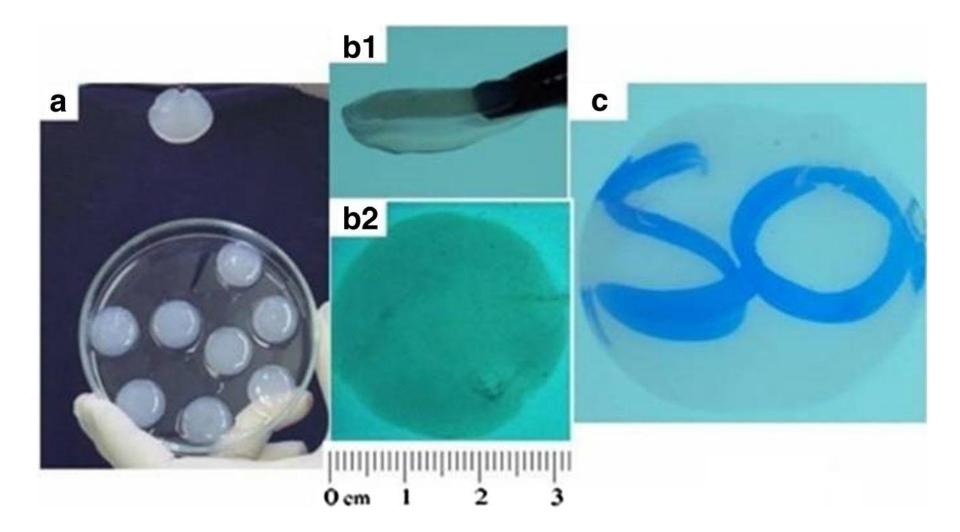
Appl Microbiol Biotechnol

Table 1 Conditions, yields, production methods and microorganisms used to produce bacterial nanocellulose described in the literature starting in 2006

| Sucrose 3.83 Static culture at 30 °C for 96 h G. xylinus ATCC 53524 (Mikkelsen et al. 2009) Acetobacter xylinum NBRC 13693 Glucose in the presence of MCP-1 1.20 Stirred culture at 30 °C and 125 rpm for 288 h Molasses and corn steep liquor in the presence of acetic acid Glucose 3.12 Stirred culture at 30 °C and 200 rpm for 168 h Glucose 4. Acetobacter xylinum (Hu and Catchmark 2010) JCM 9730 2010) Jung et al. 2010) Static culture at 30 °C for 96 h G. sacchari (Trovatti et al. 2011) Glucose (HS broth supplemented with n- butanol) HS broth supplemented with thin stillage 10.22 Static culture at 30 °C for 168 h G. sacchari Gomes et al. (Gomes al. 2013) | Carbon source | BNC Yield (g/L) | Production method | Microorganism | Reference | |
|--|---|--------------------|------------------------------------|-------------------------|------------------------------------|--|
| Orange juice containing nitrogen sources of 5.90 HS Glucose in the presence of MCP-1 1.20 Stirred culture at 30 °C and 125 rpm for 288 h JCM 9730 2010) Molasses and corn steep liquor in the presence of acetic acid Glucose 2.70 Static culture at 30 °C and 200 rpm for 168 h Static culture at 30 °C for 96 h Static culture at 30 °C and 200 rpm for 168 h Glucose (HS broth supplemented with n-butanol) HS broth supplemented with thin stillage 10.22 Static culture at 30 °C for 144 h A. xylinum 186 (Lu et al. 2011) Molasses 1.64 Static semicontinuous process for 168 h Waste beer yeast treated with ultrasonication 7.02 Stirred culture at 30 °C for 168 h Static culture at 30 °C for 168 h G. xylinus (BCRC 12334) (Wu and Liu 2012) Gomes et al. (Gomes al. 2013) (Wu and Liu 2012) Static semicontinuous process for 168 h Static semicontinuous process for 168 h Waste beer yeast treated with ultrasonication 7.02 Stirred culture at 30 °C and 168 h Static culture at 30 °C and 150 rpm 3917 2014) Rotten fruit culture 60 Static culture at 30 °C and 96 h G. xylinus (FCO1) Static culture at 30 °C and 96 h G. xylinus ATCC 53582 (Jozala et al. 2015) Wood hot water extract 0.15 Static culture at 30 °C for 144 h G. xylinus CGMCC 2955 (Li et al. 2015) Waste water of candied jujube hydrolysate 2.25 Static culture at 30 °C for 144 h G. xylinus CGMCC 2955 (Li et al. 2015) | HS in the presence of 1 % lignosulfonate | 16.32 | Static culture at 28 °C for 168 h | G. xylinus IFO 13693 | (Keshk and Sameshima 2006) | |
| HS Glucose in the presence of MCP-1 1.20 Stirred culture at 30 °C and 125 rpm for 288 h JCM 9730 2010) Molasses and corn steep liquor in the presence of acetic acid Glucose 2.70 Static culture at 30 °C for 96 h Static culture at 30 °C for 144 h A. xylinum 186 (Lu et al. 2011) HS broth supplemented with thin stillage 10.22 Static culture at 30 °C for 168 h G. sacchari (Trovatti et al. 2011) HS broth supplemented with thin stillage 10.22 Static culture at 30 °C for 168 h G. sacchari Gomes et al. (Gomes al. 2013) Molasses 1.64 Static semicontinuous process for 168 h Static semicontinuous process for 168 h Static culture at 30 °C and 2014) Waste beer yeast treated with ultrasonication 7.02 Stirred culture at 30 °C for 168 h G. sacchari Gomes et al. (Gomes al. 2013) Gomes et al. (Gomes al. 2013) Calcar et 2014) Static culture at 30 °C and 3917 Stirred culture at 30 °C and 96 h G. sacchari Gomes et al. (Cakar et 2014) Static culture at 30 °C and 96 h Static culture at 30 °C and 96 h G. sacchari Gomes et al. (Cakar et 2014) Static culture at 30 °C and 96 h Static culture at 30 °C and 96 h G. sacchari Gomes et al. (Cakar et 2014) Calcar et 2014) Static culture at 30 °C and 96 h G. sacchari Gomes et al. (Cakar et 2014) Static culture at 30 °C and 96 h G. sacchari Gomes et al. (Cakar et 2015) Static culture at 30 °C and 96 h G. sacchari Gomes et al. (Cakar et 2014) Calcar et 2014) Static culture at 30 °C and 96 h G. sacchari Gomes et al. (Cakar et 2015) Static culture at 30 °C for 144 h G. sylinus GGMCC 2955 (Li et al. 2015) Waste water of candied jujube hydrolysate 2.25 Static culture at 30 °C for 144 h G. sylinus CGMCC 2955 (Li et al. 2015) | Sucrose | 3.83 | Static culture at 30 °C for 96 h | G. xylinus ATCC 53524 | (Mikkelsen et al. 2009) | |
| Molasses and com steep liquor in the presence of acetic acid Glucose 2.70 Static culture at 30 °C for 96 h By broth supplemented with n-butanol) HS broth supplemented with thin stillage 10.22 Static culture at 30 °C for 168 h Static culture at 30 °C for 168 h G. sacchari (Trovatti et al. 2011)) Molasses 1.64 Static culture at 30 °C for 168 h G. sacchari Gomes et al. (Gomes al. 2013) Molasses 1.64 Static semicontinuous process for 168 h Waste beer yeast treated with ultrasonication 7.02 Static culture at 30 °C and 168 h Static semicontinuous process for 168 h Static culture at 30 °C and 168 h G. sacchari Gomes et al. (Gomes al. 2013) G. sacchari Gomes et al. (Gomes al. 2013) G. sacchari Gomes et al. (Cakar et 168 h Static semicontinuous process for 168 h Static culture at 30 °C and 168 h G. sacchari Gomes et al. (Cakar et 168 h Static semicontinuous process for 168 h Static semicontinuous process for 168 h Static culture at 30 °C and 168 h G. sacchari Gomes et al. (Cakar et 2014) Cakar et al. (Cakar et 2014) Stirred culture at 30 °C and 96 h Static culture at 30 °C and 96 h G. sylinus (FCO1) Cakar et al. (Lin et al. (Lin et al. 2015) Static culture at 30 °C and 96 h G. sylinus ATCC 53582 Jozala et al. 2015) Waste water of candied jujube hydrolysate 2.25 Static culture at 30 °C for 144 h G. sylinus CGMCC 2955 (Li et al. 2015) Citrus Juice and sucrose - Static culture at 30 °C G. sp. gel_SEA623-2 (Kim et al. 2015) | | 5.90 | Static culture at 30 °C for 96 h | | (Kurosumi et al. 2009) | |
| presence of acetic acid Glucose 2.70 Static culture at 30 °C for 96 h G. sacchari (Trovatti et al. 2011) Glucose (HS broth supplemented with n- butanol) HS broth supplemented with thin stillage 10.22 Static culture at 30 °C for 144 h A. xylinum 186 (Lu et al. 2011) HS broth supplemented with thin stillage 10.22 Static culture at 30 °C for 168 h G. xylinus (BCRC 12334) (Wu and Liu 2012) G. sacchari Gomes et al. (Gomes al. 2013) Molasses 1.64 Static semicontinuous process for 168 h Catheria and 2014) Waste beer yeast treated with ultrasonication 7.02 Stirred culture at 30 °C and 150 rpm 3917 Rotten fruit culture 60 Static culture at 30 °C and 96 h G. xylinus ATCC 53582 (Jozala et al. 2015) Wood hot water extract 0.15 Static culture at 30 °C for 144 h Catheria and 23,769 Waste water of candied jujube hydrolysate 2.25 Static culture at 30 °C for 144 h G. xylinus CGMCC 2955 (Kim et al. 2015) Static culture at 30 °C G. sp. gel_SEA623-2 (Kim et al. 2015) | Glucose in the presence of MCP-1 | 1.20 | | | (| |
| Glucose (HS broth supplemented with n-butanol) HS broth supplemented with thin stillage 10.22 Static culture at 30 °C for 144 h A. xylinum 186 (Lu et al. 2011) HS broth supplemented with thin stillage 10.22 Industrial residues from olive oil production 1.28 - G. sacchari Gomes et al. (Gomes al. 2013) Molasses 1.64 Static semicontinuous process for 168 h Waste beer yeast treated with ultrasonication 7.02 Stirred culture at 30 °C and 150 rpm 3917 Rotten fruit culture 60 Static culture at 30 °C and 96 h G. xylinus (FC01) Static culture at 30 °C and 96 h G. xylinus ATCC 53582 (Jozala et al. 2015) Wood hot water extract 0.15 Static culture at 28 °C for 672 h Acetobacter xylinum 23,769 2015b) Waste water of candied jujube hydrolysate 2.25 Static culture at 30 °C for 144 h G. xylinus CGMCC 2955 (Li et al. 2015) Citrus Juice and sucrose - Static culture at 30 °C G. sp. gel_SEA623-2 (Kim et al. 2015) | | 3.12 | | Acetobacter sp. V6 | (Jung et al. 2010)) | |
| Static culture at 30 °C for 168 h G. xylimus (BCRC 12334) (Wu and Liu 2012) | Glucose | 2.70 | Static culture at 30 °C for 96 h | G. sacchari | (Trovatti et al. 2011)) | |
| Industrial residues from olive oil production 1.28 - G. sacchari Gomes et al. (Gomes al. 2013) Molasses 1.64 Static semicontinuous process for 168 h Static culture at 30 °C and 169 h Static culture at 30 °C and 96 h Gomes et al. (Gomes al. 2013) Gakar et al. (Cakar et 2014) Lin et al. (Lin et al. 2014) Static culture at 30 °C and 96 h Gomes et al. (Gomes al. 2013) Gakar et al. (Cakar et 2014) Static culture at 30 °C and 96 h Gomes et al. (Gomes al. 2014) Static culture at 30 °C and Gomes at al. 2015) Gakar et al. (Cakar et 2014) Static culture at 30 °C and 96 h Gomes et al. (Gomes al. 2014) Static culture at 30 °C and Gomes al. 2014 Gomes et al. (Gomes al. 2014) Static culture at 30 °C and Gomes at al. 2015 Gomes et al. (Gomes al. 2013) Gomes et al. (Gomes al. 2013) Static culture at 30 °C and Gomes al. 2015 Gomes et al. (Gomes al. 2013) Gomes et al. (Gomes al. 2014) Gomes al. 2013) | | 1.33 | Static culture at 30 °C for 144 h | A. xylinum 186 | (Lu et al. 2011) | |
| Molasses 1.64 Static semicontinuous process for 168 h Waste beer yeast treated with ultrasonication 7.02 Stirred culture at 30 °C and 150 rpm Rotten fruit culture 60 Static culture at 30 °C and 96 h Wood hot water extract 0.15 Static culture at 28 °C for 672 h Waste water of candied jujube hydrolysate 2.25 Static culture at 30 °C for 144 h Citrus Juice and sucrose 31. 2013) Cakar et al. (Cakar et 2014) Cakar et 2014 Cakar et 2015 Cakar et | HS broth supplemented with thin stillage | 10.22 | Static culture at 30 °C for 168 h | G. xylinus (BCRC 12334) | (Wu and Liu 2012) | |
| Waste beer yeast treated with ultrasonication 7.02 Stirred culture at 30 °C and 150 rpm Rotten fruit culture 60 Static culture at 30 °C and 96 h Wood hot water extract 0.15 Static culture at 28 °C for 672 h Waste water of candied jujube hydrolysate 2.25 Static culture at 30 °C for 144 h Citrus Juice and sucrose 168 h 2014) Cin et al. (Lin et al. 2015) G. kansenii CGMCC 3917 Citrus Juice at 30 °C and 96 h Acetobacter xylinum 23,769 Citrus Juice and sucrose Citrus Juice and sucrose Static culture at 30 °C for 144 h G. xylinus CGMCC 2955 Citrus Juice and sucrose Kim et al. 2015) | Industrial residues from olive oil production | 1.28 | - | G. sacchari | Gomes et al. (Gomes e al. 2013) | |
| Rotten fruit culture 60 Static culture at 30 °C and 96 h G. xylinus ATCC 53582 (Jozala et al. 2015)) Wood hot water extract 0.15 Static culture at 28 °C for 672 h Acetobacter xylinum (Erbas Kiziltas et al. 2015b) Waste water of candied jujube hydrolysate 2.25 Static culture at 30 °C for 144 h G. xylinus CGMCC 2955 (Li et al. 2015) Citrus Juice and sucrose – Static culture at 30 °C G. sp. gel_SEA623-2 (Kim et al. 2015) | Molasses | 1.64 | - | G. xylinus (FC01) | Çakar et al. (Cakar et a 2014) | |
| Wood hot water extract 0.15 Static culture at 28 °C for 672 h Acetobacter xylinum 23,769 Citrus Juice and sucrose 0.15 Static culture at 28 °C for 672 h Acetobacter xylinum 23,769 Citrus Juice and sucrose | Waste beer yeast treated with ultrasonication | 7.02 | | | | |
| Waste water of candied jujube hydrolysate 2.25 Static culture at 30 °C for 144 h G. xylinus CGMCC 2955 (Li et al. 2015) Citrus Juice and sucrose – Static culture at 30 °C G. sp. gel_SEA623-2 (Kim et al. 2015) | Rotten fruit culture | 60 | Static culture at 30 °C and 96 h | G. xylinus ATCC 53582 | (Jozala et al. 2015)) | |
| Citrus Juice and sucrose – Static culture at 30 °C G. sp. gel_SEA623-2 (Kim et al. 2015) | Wood hot water extract | 0.15 | Static culture at 28 °C for 672 h | _ | | |
| | Waste water of candied jujube hydrolysate | 2.25 | Static culture at 30 °C for 144 h | G. xylinus CGMCC 2955 | (Li et al. 2015) | |
| Lipid fermentation wastewater 0.66 Static culture at 28 °C for 5 days G. xylinus CH001 (Huang et al. 2016) | Citrus Juice and sucrose | _ | Static culture at 30 °C | G. sp. gel_SEA623-2 | (Kim et al. 2015) | |
| | Lipid fermentation wastewater | 0.66 | Static culture at 28 °C for 5 days | G. xylinus CH001 | (Huang et al. 2016) | |

Jozala et al, Appl Microbiol Biotechnol. 2016, 100(5):2063-72

Bacterial Nanocellulose produced by culture medium composed by rotten fruits: A) After treatment with NaOH; B1, B2 size and thickness, C transparency.



Bacterial nanocellulose applications in different areas

| Area | Application |
|-------------------------|---|
| Cosmetics | Stabilizer of emulsions like creams, tonics, conditioners, nail polishes. |
| Textile industry | Sports clothing, tents and camping equipment |
| Mining and refinery | Sponges to collect leaking oil, materials for absorbing toxins. |
| Waste treatment | Recycling of minerals and oils |
| Sewage purification | Urban sewage purification, ultra filtration water |
| Communications | Diaphragms for microphones and stereo headphones |
| Food industry | Edible cellulose (nata de coco) |
| Paper industry | Artificial replacement of wood, special papers |
| Medicine/ biomedical | Temporary artificial skin for burns and ulcers, dental implant components; Antimicrobial wound dressing, Nanofilm, Drug Delivery, Drug excipient. |
| Laboratories | Protein immobilization, chromatographic techniques, tissue culture medium |
| Electronics | Opto-electronics materials (liquid crystal displays) |
| Energy | Membrane fuel cell (palladium) |

Pesticides



NATURAL PESTICIDES

- > Secondary metabolites of microorganisms, plants and insects
- > Spinosad insecticide of soil actinomycete (Saccharopolyspora spinosa)
 - Bacillus thuringiensis pesticide
 - Insect pheromones...

Biofuels

Fuels - Biodiesel and Bioethanol

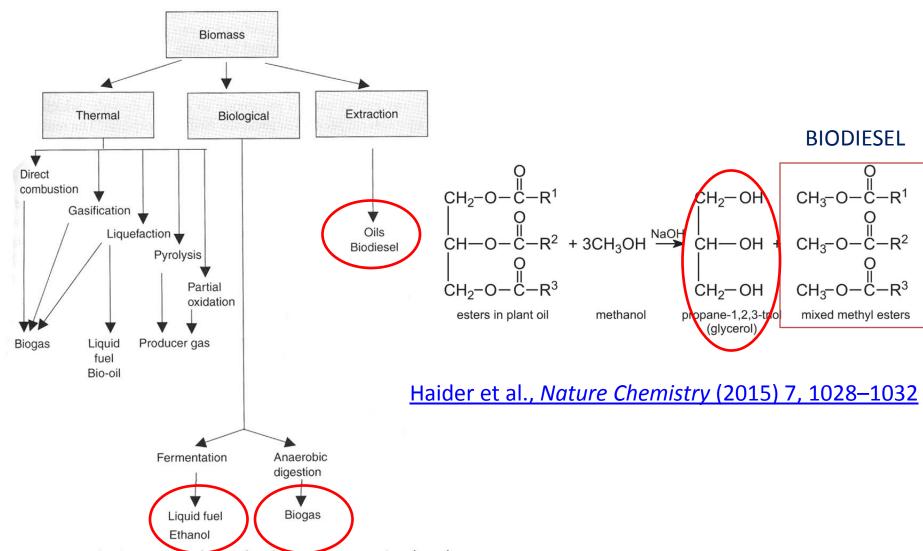


Figure 7.17 The routes for the extraction of energy from biomass using extraction, thermal, and biological methods.

Metal nanoparticles

Recently, synthesizing metal nanoparticles using microorganisms and plants has been extensively studied and has been recognized as a green and efficient way for further exploiting microorganisms as convenient nanofactories!

agricultural, environmental, and physiochemical areas.

Applications of metal nanoparticles: electrochemical sensors and biosensors, biomedical,

Gold nanoparticles: specific delivery of drugs, for tumor detection, angiogenesis, genetic disease and genetic disorder diagnosis, photoimaging, and photothermal therapy.

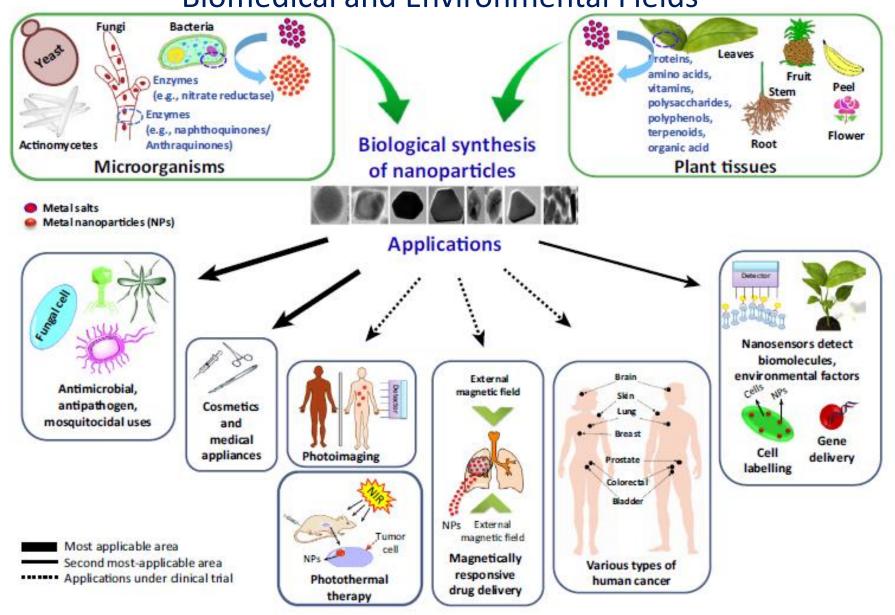
Iron oxide nanoparticles: for cancer therapy, hyperthermia, drug delivery, tissue repair, cell labeling, targeting and immunoassays, detoxification of biological fluids, magnetic resonance imaging, and magnetically responsive drug delivery therapy.

Silver nanoparticles: antimicrobial purposes, anticancer, anti-inflammatory, and wound treatment applications.

Zinc and **titanium** nanoparticles: biomedicine, cosmetic, ultraviolet (UV)-blocking agents, and various cutting-edge processing applications.

Copper and **palladium** nanoparticles: batteries, polymers, plastics plasmonic wave guides, and optical limiting devices, as antimicrobial agents.

Biological Synthesis and Applications of Metal Nanoparticles in Biomedical and Environmental Fields

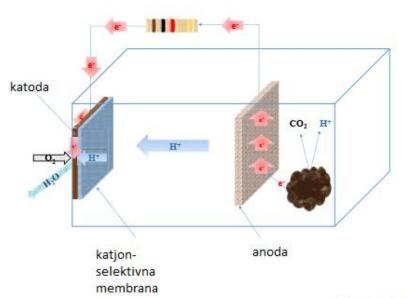


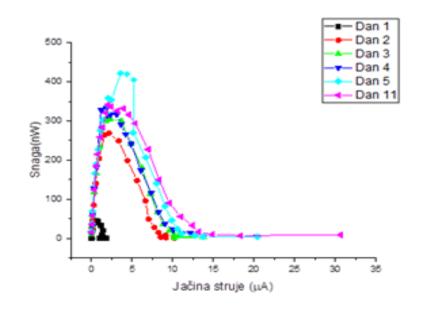
Singh et al., Trends in Biotechnology, July 2016, Vol. 34, No.

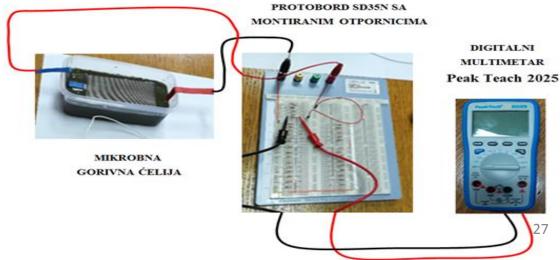
Microbial Fuel Cells

Electricity from Microorganisms

Grafik zavisnosti snage razvijene na nizu otpornika od jačine struje koja protiče kroz otpornike

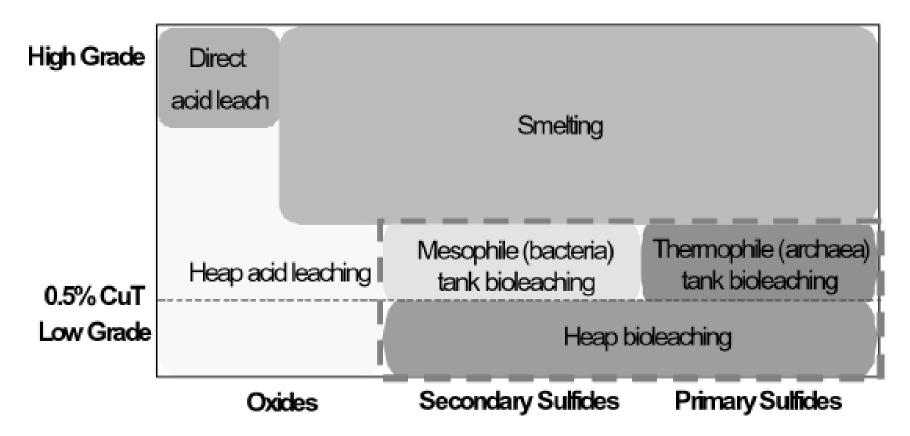






Biohydrometallurgy

Copper Extraction Technologies



Hydrometallurgical:

Mining, crushing, milling, flotation => Copper concentrate => smelter proces => metal

Biohydrometallurgical:

Mining, crushing, milling, => acidic sulfate solution => solvent extraction or electrowinning

BASICS IN BACTERIAL METAL LEACHING IN THE PRESENCE OF PYRITE (FeS₂)

$$MS_{(s)} + 2O_{2} \longrightarrow M^{2+} + SO_{4}^{2-} \qquad At.spp. \quad (B)$$

$$FeS_{2(s)} + 3,5O_{2} + 0,5H_{2}O \longrightarrow Fe^{3+} + H^{+} + 2SO_{4}^{2-} \qquad At.f. \quad (B)$$

$$2MS_{2(s)} + 10Fe^{3+} + 4H_{2}O \longrightarrow 2M^{2+} + 10Fe^{2+} + S + 8H^{+} + SO_{4}^{2-} \qquad (H)$$

$$FeS_{2(s)} + 2Fe^{3+} \longrightarrow 3Fe^{2+} + 2S \qquad (H)$$

$$S + 6Fe^{3+} + 4H_{2}O \longrightarrow 6Fe^{2+} + 8H^{+} + SO_{4}^{2-} \qquad (H)$$

$$MS_{(s)} \longrightarrow M^{2+} + S + 2e^{-} \qquad (E)$$

$$0,5O_{2} + 2H^{+} \longrightarrow H_{2}O - 2e^{-} \qquad (E)$$

$$Fe^{2+} + 0,2SO_{2} + H^{+} \longrightarrow Fe^{3+} + 0,5H_{2}O \qquad At.f. \quad (B)$$

$$S + 1,5O_{2} + H_{2}O \longrightarrow 2H^{+} + SO_{4}^{2-} \qquad At.th. \quad (B)$$

$$Fe^{3+} + 3H_{2}O \longrightarrow Fe(OH)_{3(s)} + 3H^{+} \qquad (H)$$

$$M(O,CO_{3},SiO_{3})_{(s)} + 2H^{+} \longrightarrow M^{2+} + (CO_{2},SiO_{2}) + H_{2}O \qquad (H)$$

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HEAP LEACHING PLANT FOR EXTRACTION OF COPPER FROM OXIDIC ORE IN LOMAS BAYAS, CHILE



Pad-drainage system



Irrigated heap drippers



Stacker



Copper leachate from heap to pond

GEOCOAT® PLANT, EXTRACTION OF GOLD, SOUTH AFRICA

1893-2003

❖INOCULUM BUILDUP (Acidithiobacillus ferrooxidans, Acidithiobacillus thiooxidans, Leptospirillum ferrooxidans, thermophilic archaea Sulfolobus and Acidianus)



BIOX® plant, extraction of **GOLD**, South Africa





Mešana populacija: Acidithiobacillus ferrooxidans, Acidithiobacillus thiooxidans, Leptospirillum ferrooxidans, Acidithiobacillus caldus, Leptospirillum ferriohilum, Ferroplasma-like archaea

| | | Monthly Average | | | | | | |
|-----------------------------------|-----|-----------------|--------|--------|--------|--------|--------|--|
| | | May-05 | Jun-05 | Jul-05 | Aug-05 | Sep-05 | Oct-05 | |
| Concentrate treated | tpd | 64.1 | 66.3 | 69.2 | 62.2 | 65.6 | 69.7 | |
| Concentrate gold grade | g/t | 103.6 | 118.0 | 137.1 | 91.9 | 114.0 | 102.9 | |
| Concentrate S ²⁻ grade | % | 14.3 | 15.3 | 14.2 | 14.2 | 15.2 | 14.4 | |
| Sulphide oxidation | % | 95.9 | 95.8 | 95.2 | 95.3 | 95.9 | 96.1 | |
| Gold recovery | % | 97.5 | 96.7 | 96.9 | 96.8 | 97.3 | 97.2 | |
| BIOX® availability | % | 100.0 | 99.6 | 98.8 | 98.8 | 97.4 | 99.4 | |

ACID MINE DRAINAGE

- 1. Mineral processing and other anthropogenic activities leave by-products exposed to oxygen and moisture in the air. If the by-products are **sulfide minerals** they may start to **bioleach** out of control and cause the environmental problem known as acid mine drainage (AMD). AMD manifests itself as acidified groundwater contaminated by heavy metals.
- 2. Drainage flowing from or caused by surface mining or coal refuse piles that are typically **highly acidic** with elevated levels of **dissolved metals**.

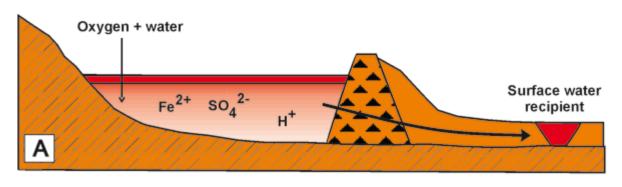
Acid mine drainage is formed by uncontrolled bioleaching reactions
- a series of complex geochemical and microbial reactions that occur when water and oxygen come into contact with pyrite (iron disulfide minerals) in for example coal, refuse or the overburden of a mine operation.

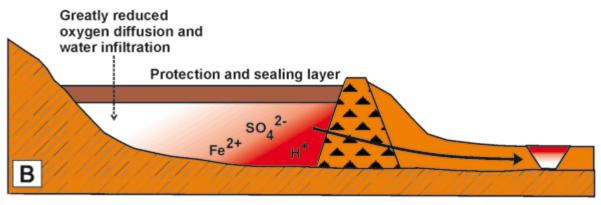


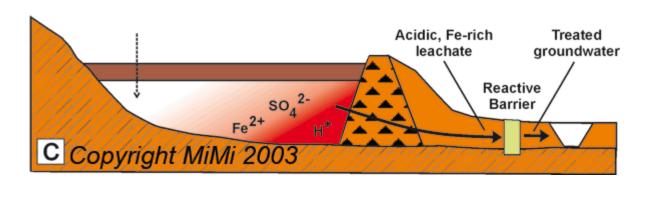
ACID MINE DRAINAGE

Copper mine Bor



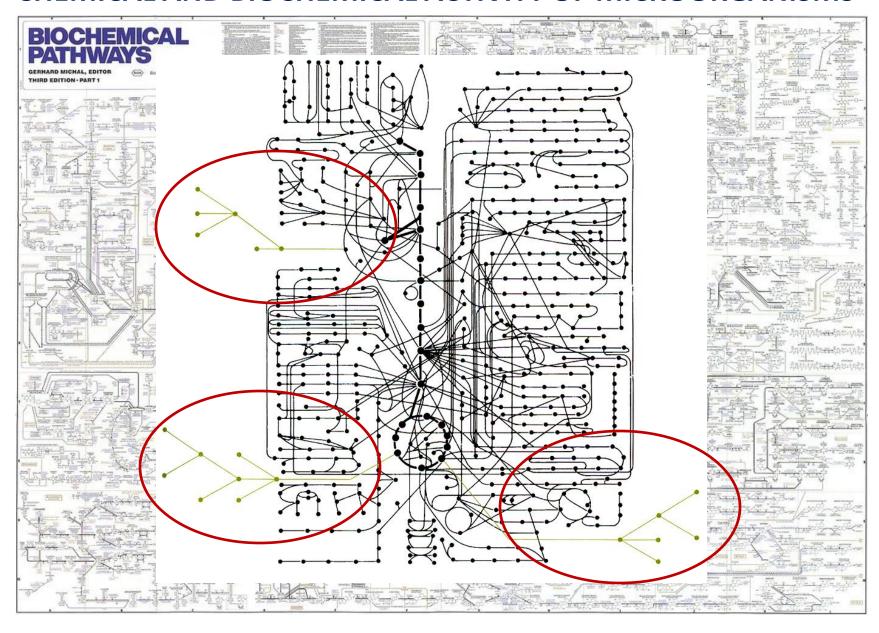






Bioremediation

CHEMICAL AND BIOCHEMICAL ACTIVITY OF MICROORGANISMS



Thank you for your attention